

El-Hayah

JURNAL BIOLOGI

Journal Homepage: <http://ejournal.uin-malang.ac.id/index.php/bio/index>
e-ISSN: 2460-7207, p-ISSN: 2086-0064

Original research article

Contribution of Drosophila Climbing Assay in Studying Biology and Diseases

Ahmad Fauzi¹, Muhamad Justitia Ramadhan², Natasya Adiba Zahrah², Sinta Kharomah², Hikmah Buroidah², Nindiana Choirunisa^{1,2}, Hidayati Maghfiroh², Siti Zubaidah^{2*}

¹ Universitas Muhammadiyah Malang, Jl. Raya Tlogomas 246 Malang 65144

² Universitas Negeri Malang, Jl. Semarang No. 5, Malang 65145

*Corresponding author

Email: siti.zubaidah.fmipa@um.ac.id

DOI: [10.18860/elha.v9i4.24210](https://doi.org/10.18860/elha.v9i4.24210)

Article Info

Article history:

Received 30 November 2023

Received in revised form 29

January 2024

Accepted 23 March 2024

Key Word:

Climbing assay *Drosophila melanogaster* Systematic literature review

Abstract

The *Drosophila Climbing Assay* (DCA) is an essential assay for disease and other biological studies. This systematic literature review (SLR) aims to analyze DCA's distribution, treatment, and contribution during the 21st Century. VOS Viewer was also involved in analyzing the bibliometric and gap analysis in related studies. By using the Scopus database and limiting document types to journal articles, a total of 183 documents have been successfully collected in this SLR. After going through a selection process based on inclusion and exclusion criteria and eligibility assessment through the PRISMA procedure, 163 articles remain that can be analyzed. Behaviour, model, activity, expression, lifespan, and disease are terms that often equate to studies involving the DCA procedure. The U.S., China, and India are the three countries that most frequently report DCA. DCA has been involved intensively in behaviour, Parkinson's, and nervous system research. The co-occurrence analysis resulted in 5 clusters, and the DCA procedure to study the impact of nutrient stress and nanoparticles resulted from the identified gap analysis.

1. INTRODUCTION

Drosophila melanogaster, or the fruit fly, has become pivotal in advancing biological science [1], [2]. Its small size, short life cycle, high reproductive rate, and ease of observing traits in this organism have made *D. melanogaster* a favoured subject in biological experiments [3]. These insects have helped

researchers study various concepts in the field of genetics [4]–[6] and development [7]. This model organism has also been involved in uncovering various complex biological phenomena in physiology [8] and neurobiology [9].

Until the last decade, *Drosophila* has continued to establish itself as one of the most popular model organisms in various

laboratories in many countries [1], [2], [4]–[6]. Along with the development of bioinformatics and molecular biology techniques, researchers can condition *Drosophila* to become an organism capable of modelling various biological conditions [7], [10], [11]. Moreover, because its genome is similar to the human genome, this insect has significantly contributed to studying various human diseases [4]. To study various biological phenomena and diseases, researchers have developed various assays whose data can be used as essential indicators of various specific biological conditions.

One of the assays that was developed and is often involved in research involving *Drosophila* is the *Drosophila* Climbing Assay (DCA) [12], [13]. This assay involves placing the flies in a vial and observing their ability to climb upward against gravity. Despite its simplicity, this assay informs about the locomotor function of the tested *Drosophila* and provides data related to their behaviour. Furthermore, this assay can serve as an indicator of the condition of the *Drosophila* nervous system. Given its cost-effectiveness and simplicity, along with its significance as a vital health indicator, the DCA has been incorporated into various biological research studies up to the present day.

Although the DCA has been extensively documented in various scientific publications, these reports primarily consist of laboratory-based experimental research and methodological development related to the DCA. On the other hand, comprehensive

reviews assessing the utilization of DCA up to the present time are scarce. Given the multitude of studies incorporating DCA into their research designs, there is a compelling need for a systematic literature review (SLR) focused on using this assay. Therefore, this paper initiates an SLR specifically targeting the DCA. This SLR aims to provide a holistic perspective on the application, variations, and outcomes of DCA testing in studies conducted across different countries. It has the potential to unearth existing gaps, facilitate discussions concerning standardized protocols that researchers can adopt, and guide the direction of future DCA research.

2. MATERIALS AND METHODS

This SLR adheres to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines, the process of which is outlined in Figure 1. The primary research question posed in this SLR is, "How does the *Drosophila* Climbing Assay (DCA) contribute to the study of various biological conditions and diseases?" The selected database for this study is Scopus. All publications gathered encompass studies in which DCA is utilized as one of its data sources. The search query employed is presented in Table 1, and the inclusion and exclusion criteria are detailed in the same table. After collecting the relevant papers, the screening process commences by selecting papers based on the predefined inclusion and exclusion criteria (Table 1).

Table 1. Search query, inclusion criteria, and exclusion criteria used

Components	Description
Search query	TITLE-ABS-KEY ("climbing ability" OR "climbing behaviour" OR "climbing assay" OR "negative geotaxis" OR "wall climbing") AND ALL ("drosophila" OR "fruit fly") AND ALL ("imago" OR "adult")
Inclusion criteria	articles that were published before 2023, classified as journal articles, in the final publication stage, sourced from journals, written in English, and available as open access.
Exclusion criteria	paper reviews, full papers that were inaccessible, non-original article

Based on Figure 1, 331 publication titles emerged as search results using the search query provided in Table 1. Since no duplicate papers were identified, all papers proceeded to the screening stage. During this stage, papers published before 2023, categorized as journal articles, in the final publication stage, sourced from scientific journals, written in English, and open access in nature were retained, resulting in 183 titles. Furthermore, five full papers remained inaccessible, leaving 176 papers eligible for evaluation. Among these, five papers merely demonstrated or introduced methods, assays, or tools, and eight papers did not involve *Drosophila* as the subject of their research. Consequently, 163 papers remain for analysis in this systematic literature review (SLR).

Data extraction was conducted on the remaining 163 papers during the data analysis phase. Based on the research questions posed in this systematic literature review (SLR), several key points were scrutinized in each paper, including the positioning of *Drosophila* as a model organism, the biological topic underlying the research problem, and the specific DCA techniques designed by the researchers. Data analysis was performed using a qualitative approach, and the results served as the foundation for data synthesis. Additionally, a bibliometric analysis was conducted to elucidate the relationships between concepts across all the collected publications.

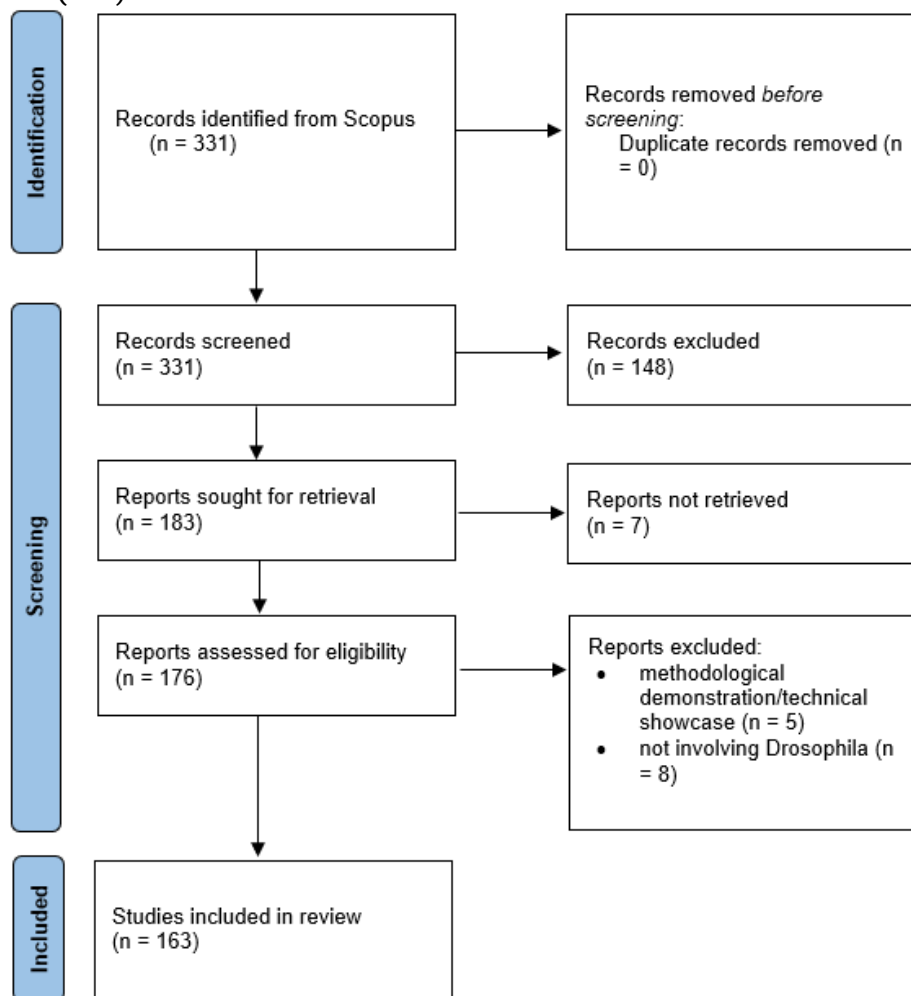


Figure 1. PRISMA steps in this SLR study

3. Results and discussion

Based on the PRISMA-based selection process, out of the 183 papers obtained from the Scopus database, 163 papers remained eligible for further analysis. The first theme used for data extraction in this SLR pertains to the role of *Drosophila* as a model organism

in the reviewed research. The results of this extraction are presented in Table 2. As indicated in Table 2, it is evident that *Drosophila* has been the subject of research modelling various conditions, spanning from modelling diverse diseases to exposure-related conditions and ageing processes.

Table 2. Data extraction for the model organism theme

Model Organism	Reference
Disease	Parkinson's Disease [14], [15], [24]–[33], [16], [34]–[38], [17]–[23], Alzheimer's Disease [39], [40], [49]–[54], [41]–[48], Huntington's disease [55]–[57], neurodegenerative disease (general) [58]–[60], Autism [61], Barth syndrome [62], Amyotrophic lateral sclerosis [63], Creutzfeldt-Jakob Disease [64], diastolic cardiac defects [65], galactosemia [66], Gerstmann–Straussler–Scheinker syndrome [11], Hereditary spastic paraplegias [55], Myotonic Dystrophy [67], spinocerebellar ataxias [68], FARS2 deficiency [7], Sepsis [69], myopathy [70], diabetes [71]
Medical condition	Brain function [72], depression [73], Mitochondrial diseases [74], gene mutation [75]–[78], neurodegeneration [79], retinal degeneration [80], NT5C2 knockdown [81], Obesity [82], Oxidative stress condition [83], sensorineural hearing loss [84], Traumatic injuries [85]–[88], Obesity [71], Autophagy down expression [89], other [90]–[92]
Ageing	age-dependent neurodegeneration [93], age-related behaviour [94], age-related locomotor impairment [95], age-related muscle [10], aging (general) [96]–[100]
Substance exposure	acute exposure to ethanol [101], nanoparticle exposure [102]–[107], Chemotherapy [108], [109], stimulant drug consumption [110], toxicity and heavy metal exposure [111]–[114], herbicide exposure [115], [116], pest [117], pollutant exposure [118], [119], quercetin consumption [120]
Alternative medicine consumption	medical plant substance/traditional medicine evaluation [121]–[125], other [126]
Diet	Dietary restriction [127], [128], Dietary supplementation [129], dietary protein [130], AKG consumption [131], High Fat Diet [132]–[136], high-salt diet [137], MSG consumption [138], non-caloric artificial sweeteners consumption [139], probiotic expose [140]
Physical activity	Exercise training [141]–[144]
Physical condition	Electromagnetic/geomagnetic field effect [145], [146], hypergravity [147], radiation exposure [148], [149], oxygen deprivation [150]
Social condition	male presence [151], predation exposure [152], Social space variation [153]
Other	Other conditions [5], [6], [162]–[171], [154]–[161]

Drosophila is capable of modelling various human diseases primarily due to the high degree of genetic conservation between this organism and humans [172], [173]. Numerous fundamental cellular and molecular pathways are highly conserved in *Drosophila*, enabling researchers to investigate various human diseases' genetic and physiological mechanisms [174], [175]. This genetic similarity, combined with the simplicity and well-characterized genome of *Drosophila*, has facilitated researchers in identifying and manipulating specific genes relevant to various human disease conditions. Additionally, developing various molecular techniques, such as gene knockout, overexpression, and downregulation, has allowed researchers to condition *Drosophila* to model the health conditions under investigation [176].

In addition to modelling diseases, the short lifespan of *Drosophila* makes it easy for researchers to study the aging process in this organism [93], [94]. Researchers also easily observe various processes and conditions related to aging more quickly than other model organisms. Apart from that, because the generation time is short, but the number of derivatives is significant, researchers will be more efficient in carrying out drug testing on genetic screens on a large scale.

Based on Table 2, the DCA has also been involved in studies positioning *Drosophila* to model the effects of physical activity [141]–[144], physical factors [145]–[147], social factors [151]–[153], and dietary conditions [132]–[136]. *Drosophila* has innate locomotor abilities that are needed in studies where treatment involves physical activity. Apart from that, *Drosophila* also has various complex behaviours that certain physical activities can influence. Furthermore, *Drosophila* is also sensitive to various environmental conditions, including physical factors [145]–[147]. Because of the small size of the culture, researchers can easily manipulate various physical factors in their laboratories. Furthermore, their social behaviours, including courtship, aggression,

and mating, are thoroughly documented and can be manipulated to explore the influence of social factors on gene expression, neurobiology, and overall health [177], [178]. These studies illuminate how social environments can shape biological outcomes. Lastly, *Drosophila*'s responsiveness to dietary conditions is a consequence of their dietary adaptability and the ease of dietary control in laboratory settings [127], [128], [132]–[136]. Researchers can precisely tailor the composition of their food, simulating various dietary scenarios. This adaptability facilitates investigations into the effects of diet on metabolism, aging, and susceptibility to diseases.

In the second theme, the SLR analysis was directed toward mapping the biological topics underpinning the research issues reported in the reviewed articles. Based on the extraction results, the *Drosophila* Climbing Assay (DCA) was most frequently involved in studies related to the nervous system [14], [15], [24]–[33], [16], [34]–[43], [17], [44]–[53], [18], [54], [58]–[60], [19]–[23]. This climbing assay, a straightforward yet powerful behavioural test, provides researchers with valuable insights into the functional status of the nervous system and its modifications under various conditions or diseases. In this context, the primary role of the nervous system is to control motor activity. On the other hand, DCA serves as a robust indicator for assessing *Drosophila*'s motor skills.

Additionally, the DCA exhibits high sensitivity, making it suitable for detecting subtle changes in motor function. Furthermore, its non-invasive nature and capacity for high-throughput testing make it an attractive option for simultaneously assessing large groups of individuals or conditions, thereby minimizing stress on experimental subjects. Returning to Table 2 and correlating with the result of data extraction in the second theme, *Drosophila* frequently models Parkinson's Disease [14], [15], [24]–[33], [16], [34]–[38], [17]–[23] and

Alzheimer's Disease [39], [40], [49]–[54], [41]–[48], both of which are neurodegenerative diseases. The decline in negative geotaxis observed in aging flies or those modelling neurodegenerative diseases closely mirrors motor deficits seen in Parkinson's and Alzheimer's patients.

The next theme extracted through this SLR pertains to researchers' application of the DCA. The extraction results reveal that various DCA designs have been reported in the methodology sections of the analyzed articles. The wide range of climbing assay designs and data collection methods in *Drosophila* studies can be attributed to the assay's adaptability to different research questions and experimental contexts. The diversity in assay formats and collected metrics underscores the flexibility of the climbing assay in assessing various aspects of locomotor behaviour. Researchers select different assay designs and metrics based on the specific objectives of their studies. Several

studies designed a simple climbing assay with a basic procedure that involves recording the number of flies that successfully climb a specific height for a particular duration of time [11], [14], [136]. Some other studies involve more complex procedures and equipment, such as the RING assay [69], [108]. RING assay could capture nuanced aspects of climbing, such as the distance climbed or the speed of ascent, making it well-suited for studies focused on the genetic or molecular mechanisms underpinning climbing behaviour. Furthermore, the adaptability of the climbing assay lends itself to a broad spectrum of research areas beyond neurobiology, including toxicology, drug screening, and aging studies. This versatility proves particularly valuable for scientists seeking to comprehend the impacts of various factors, such as substance exposure, diet, or physical activity, on locomotor performance.

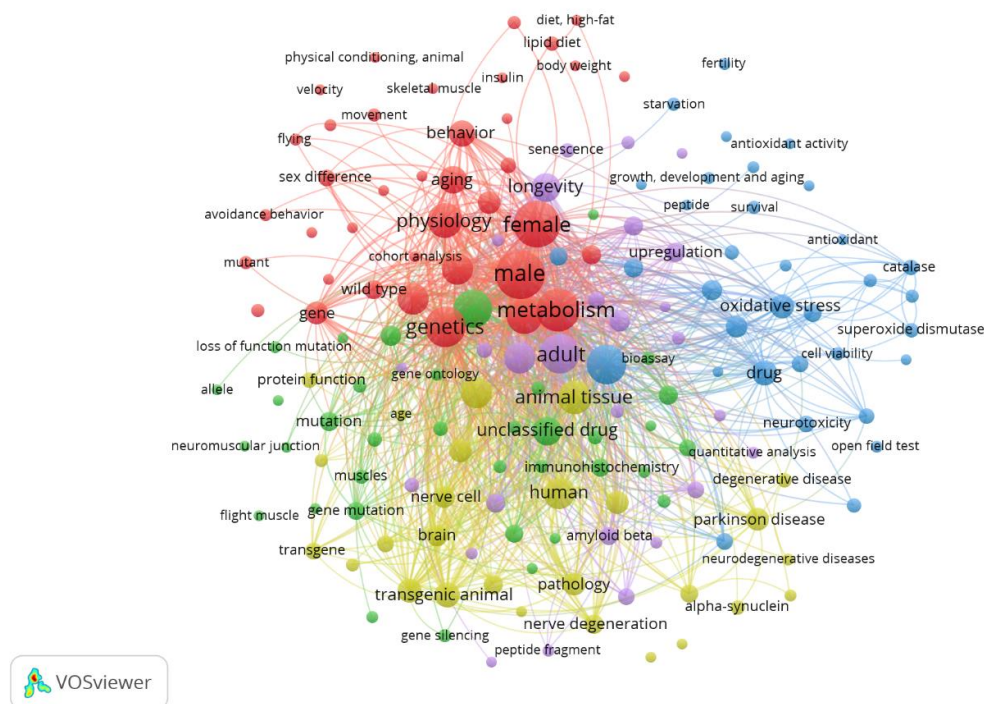


Figure 2. Results of co-occurrence analysis of index keywords

In addition to the systematic literature review (SLR), this paper also presents the

results of bibliometric analysis conducted on the reviewed papers. With the assistance of

VOS Viewer, a co-occurrence analysis of index keywords within these papers was visualized (Figure 2). "Male" and "female" emerged as the keywords with the highest occurrence values. This finding indicates that the research encompasses aspects related to gender differences in the context of *Drosophila*. It underscores the significance of understanding how gender factors influence various biological conditions and bodily responses. Keywords such as "metabolism," "genetics," "model," and "protein" also stand out, signifying that the reviewed research is closely linked to an understanding of genetic and metabolic aspects within the *Drosophila* model. Additionally, keywords such as "locomotion," "physiology," "lifespan," and "phenotype" suggest that much of the research in the context of *Drosophila* is focused on comprehending movement, physiological function, lifespan, and related phenotypes. Keywords like "transgenic animal" and "genetically modified," with a high co-occurrence count, indicate that many studies in this review may involve genetic manipulations in *Drosophila* to create models relevant to the studied conditions.

Meanwhile, keywords with lower total link strength, such as "body weight," "physical conditioning," "physiological stress," "sucrose," and "velocity," may indicate that research in the context of *Drosophila* may not explicitly encompass these aspects or may not yet have a substantial body of research to establish strong connections. The disparities in occurrence rates and total link strength between keywords reflect the research focus and trends within the reviewed *Drosophila* behavioural studies. Keywords with high occurrence rates mirror crucial aspects often serving as the primary research focus. In contrast, keywords with lower total link strength may reflect aspects that have not been fully explored or are not the primary focus of the research.

4. CONCLUSION

The systematic literature review (SLR) and bibliometric analysis offer valuable insights into the diverse research landscape surrounding *Drosophila* behaviour assays. These findings underscore the adaptability of *Drosophila* as a model organism capable of investigating a broad spectrum of topics, including neurodegenerative diseases, aging, genetic modifications, and the influence of various environmental factors. Notably, the analysis highlights keywords such as "male" and "female" with high occurrence, emphasizing the significance of considering gender differences in *Drosophila* research. Furthermore, keywords related to genetics, metabolism, and modelling take centre stage, reflecting the organism's pivotal role in genetic and metabolic studies.

Conversely, keywords with lower total link strength, such as "body weight" and "physiological stress," may suggest areas within *Drosophila* behaviour research warrant further exploration. As a suggestion for future research, prioritizing investigations into the long-term and transgenerational effects of *Drosophila* behaviour assays is recommended, as these aspects still need to be explored. Gaining insights into how behaviours and phenotypes can be inherited across generations could yield valuable information about *Drosophila*'s intricate interplay of genetic, environmental, and epigenetic factors and potentially shed light on broader biological phenomena

5. REFERENCES

- [1] Z. Mirzoyan, M. Sollazzo, M. Allocca, A. M. Valenza, D. Grifoni, and P. Bellosta, "Drosophila melanogaster: A model organism to study cancer," *Front. Genet.*, vol. 10, no. 51, pp. 1–16, Mar. 2019, doi: 10.3389/fgene.2019.00051.
- [2] M. Yamaguchi and H. Yoshida, "Drosophila as a model organism," in *Drosophila Models for Human Diseases. Advances in Experimental Medicine and*

- Biology, M. Yamaguchi, Ed. Springer, 2018, pp. 1–10.
- [3] M. D. Rand, J. M. Tennessen, T. F. C. Mackay, and R. R. H. Anholt, “Perspectives on the *Drosophila melanogaster* model for advances in toxicological science,” *Curr. Protoc.*, vol. 3, no. 8, p. e870, Aug. 2023, doi: 10.1002/cpz1.870.
- [4] F. P. Fischer, R. A. Karge, Y. G. Weber, H. Koch, S. Wolking, and A. Voigt, “*Drosophila melanogaster* as a versatile model organism to study genetic epilepsies: An overview,” *Front. Mol. Neurosci.*, vol. 16, p. 1116000, Feb. 2023, doi: 10.3389/fnmol.2023.1116000.
- [5] V. Muha et al., “O-GlcNAcase contributes to cognitive function in *Drosophila*,” *J. Biol. Chem.*, vol. 295, no. 26, pp. 8636 – 8646, 2020, doi: 10.1074/jbc.RA119.010312.
- [6] T. Cao, A. Sujkowski, T. Cobb, R. J. Wessells, and J.-P. Jin, “The glutamic acid-rich-long C-terminal extension of troponin T has a critical role in insect muscle functions,” *J. Biol. Chem.*, vol. 295, no. 12, pp. 3794 – 3807, 2020, doi: 10.1074/jbc.RA119.012014.
- [7] W. Fan et al., “FARS2 deficiency in *Drosophila* reveals the developmental delay and seizure manifested by aberrant mitochondrial tRNA metabolism,” *Nucleic Acids Res.*, vol. 49, no. 22, pp. 13108 – 13121, 2021, doi: 10.1093/nar/gkab1187.
- [8] H. Tang, L. Zhong, Y. Xu, Z. Jin, Z. Pan, and J. Shen, “Polypropylene microplastics affect the physiology in *Drosophila* model,” *Bull. Entomol. Res.*, vol. 113, no. 3, pp. 355–360, Jun. 2023, doi: 10.1017/S0007485322000633.
- [9] F. Liguori, U. B. Pandey, and F. A. Digilio, “Editorial: *Drosophila* as a model to study neurodegenerative diseases,” *Front. Neurosci.*, vol. 17, p. 1275253, 2023, doi: 10.3389/fnins.2023.1275253.
- [10] M. Ozaki, T. D. Le, and Y. H. Inoue, “Downregulating mitochondrial DNA polymerase γ in the muscle stimulated autophagy, apoptosis, and muscle aging-related phenotypes in *drosophila* adults,” *Biomolecules*, vol. 12, no. 8, 2022, doi: 10.3390/biom12081105.
- [11] J.-K. Choi et al., “A *Drosophila* model of GSS syndrome suggests defects in active zones are responsible for pathogenesis of GSS syndrome,” *Hum. Mol. Genet.*, vol. 19, no. 22, pp. 4474 – 4489, 2010, doi: 10.1093/hmg/ddq379.
- [12] A. N. Spierer, D. Yoon, C.-T. Zhu, and D. M. Rand, “FreeClimber: Automated quantification of climbing performance in *Drosophila*,” *J. Exp. Biol.*, Jan. 2020, doi: 10.1242/jeb.229377.
- [13] S. Manjila and G. Hasan, “Flight and climbing assay for assessing motor functions in *Drosophila*,” *BIO-PROTOCOL*, vol. 8, no. 5, 2018, doi: 10.21769/BioProtoc.2742.
- [14] M. C. ecili. Barone and D. Bohmann, “Assessing neurodegenerative phenotypes in *Drosophila* dopaminergic neurons by climbing assays and whole brain immunostaining,” *J. Vis. Exp.*, no. 74, p. e50339, 2013, doi: 10.3791/50339.
- [15] A. Aggarwal, H. Reichert, and K. VijayRaghavan, “A locomotor assay reveals deficits in heterozygous Parkinson’s disease model and proprioceptive mutants in adult *Drosophila*,” *Proc. Natl. Acad. Sci. U. S. A.*, vol. 116, no. 49, pp. 24830 – 24839, 2019, doi: 10.1073/pnas.1807456116.
- [16] W. Cao et al., “An automated rapid iterative negative geotaxis assay for analyzing adult climbing behavior in a *Drosophila* model of neurodegeneration,” *J. Vis. Exp.*, vol. 2017, no. 127, 2017, doi: 10.3791/56507.
- [17] L. Song et al., “Auxilin underlies progressive locomotor deficits and dopaminergic neuron loss in a *Drosophila* model of Parkinson’s disease,” *Cell Rep.*, vol. 18, no. 5, pp. 1132 – 1143, 2017, doi: 10.1016/j.celrep.2017.01.005.

- [18] P. G. M'Angale and B. E. Staveley, "Bax-inhibitor-1 knockdown phenotypes are suppressed by Buffy and exacerbate degeneration in a *Drosophila* model of Parkinson disease," *PeerJ*, vol. 5, p. e2974, 2017, doi: 10.7717/peerj.2974.
- [19] E. A. S. Musachio et al., "Bisphenol A exposure is involved in the development of Parkinson like disease in *Drosophila melanogaster*," *Food Chem. Toxicol.*, vol. 137, 2020, doi: 10.1016/j.fct.2020.111128.
- [20] J.-F. Guo et al., "Coding mutations in NUS1 contribute to Parkinson's disease," *Proc. Natl. Acad. Sci. U. S. A.*, vol. 115, no. 45, pp. 11567 – 11572, 2018, doi: 10.1073/pnas.1809969115.
- [21] Y. H. Siddique, F. Naz, Rahul, M. Rashid, and S. Mian, "Effect of Itrifal Muqawwi-e-Dimagh (a polyherbal drug) on the transgenic *Drosophila* model of Parkinson's Disease," *Phytomedicine Plus*, vol. 1, no. 4, 2021, doi: 10.1016/j.phyplu.2021.100131.
- [22] Rahul, F. Naz, S. Jyoti, and Y. H. Siddique, "Effect of kaempferol on the transgenic *Drosophila* model of Parkinson's disease," *Sci. Rep.*, vol. 10, no. 1, 2020, doi: 10.1038/s41598-020-70236-2.
- [23] Y. H. Siddique, F. Naz, Rahul, M. Rashid, and Tajuddin, "Effect of Majun Baladur on life span, climbing ability, oxidative stress and dopaminergic neurons in the transgenic *Drosophila* model of Parkinson's disease," *Heliyon*, vol. 5, no. 4, 2019, doi: 10.1016/j.heliyon.2019.e01483.
- [24] Z. Shan et al., "Effects of sevoflurane on leucine-rich repeat kinase 2-associated *Drosophila* model of Parkinson's disease," *Mol. Med. Rep.*, vol. 11, no. 3, pp. 2062 – 2070, 2015, doi: 10.3892/mmr.2014.2966.
- [25] E. J. Fernandes et al., "Exposure to lutein-loaded nanoparticles attenuates Parkinson's model-induced damage in *Drosophila melanogaster*: Restoration of dopaminergic and cholinergic system and oxidative stress indicators," *Chem. Biol. Interact.*, vol. 340, 2021, doi: 10.1016/j.cbi.2021.109431.
- [26] M. R. Poetini et al., "Hesperidin attenuates iron-induced oxidative damage and dopamine depletion in *Drosophila melanogaster* model of Parkinson's disease," *Chem. Biol. Interact.*, vol. 279, pp. 177 – 186, 2018, doi: 10.1016/j.cbi.2017.11.018.
- [27] W. Liu, K.-L. Lim, and E.-K. Tan, "Intestine-derived α -synuclein initiates and aggravates pathogenesis of Parkinson's disease in *Drosophila*," *Transl. Neurodegener.*, vol. 11, no. 1, 2022, doi: 10.1186/s40035-022-00318-w.
- [28] S. M. Pütz et al., "Loss of p21-activated kinase Mbt/PAK4 causes Parkinson-like phenotypes in *Drosophila*," *DMM Dis. Model. Mech.*, vol. 14, no. 6, 2021, doi: 10.1242/dmm.047811.
- [29] M. E. O'Hanlon, C. Tweedy, F. Scialo, R. Bass, A. Sanz, and T. K. Smulders-Srinivasan, "Mitochondrial electron transport chain defects modify Parkinson's disease phenotypes in a *Drosophila* model," *Neurobiol. Dis.*, vol. 171, p. 105803, Sep. 2022, doi: 10.1016/j.nbd.2022.105803.
- [30] S. Poddighe et al., "Mucuna pruriens (Velvet bean) rescues motor, olfactory, mitochondrial and synaptic impairment in PINK1B9 *Drosophila melanogaster* genetic model of Parkinson's disease," *PLoS One*, vol. 9, no. 10, 2014, doi: 10.1371/journal.pone.0110802.
- [31] M. A. Casu et al., "Neuroprotection by the immunomodulatory drug pomalidomide in the *Drosophila* LRRK2WD40 genetic model of parkinson's disease," *Front. Aging Neurosci.*, vol. 12, 2020, doi: 10.3389/fnagi.2020.00031.
- [32] B. T. Mannett, B. C. Capt, K. Pearman, L. M. Buhlman, J. M. Vandenbrooks, and G. B. Call, "Nicotine has a therapeutic

- window of effectiveness in a *Drosophila melanogaster* model of Parkinson's disease," *Parkinsons. Dis.*, vol. 2022, 2022, doi: 10.1155/2022/9291077.
- [33] S. K. Pirooznia et al., "PARIS induced defects in mitochondrial biogenesis drive dopamine neuron loss under conditions of parkin or PINK1 deficiency," *Mol. Neurodegener.*, vol. 15, no. 1, 2020, doi: 10.1186/s13024-020-00363-x.
- [34] Y. H. Siddique, S. Jyoti, F. Naz, and Rahul, "Protective effect of luteolin on the transgenic drosophila model of Parkinson's disease," *Brazilian J. Pharm. Sci.*, vol. 54, no. 3, 2018, doi: 10.1590/s2175-97902018000317760.
- [35] K. E. White, D. M. Humphrey, and F. Hirth, "The dopaminergic system in the aging brain of *Drosophila*," *Front. Neurosci.*, vol. 4, no. DEC, 2010, doi: 10.3389/fnins.2010.00205.
- [36] A.-R. Issa et al., "The lysosomal membrane protein LAMP2A promotes autophagic flux and prevents SNCA-induced Parkinson disease-like symptoms in the *Drosophila* brain," *Autophagy*, vol. 14, no. 11, pp. 1898 – 1910, 2018, doi: 10.1080/15548627.2018.1491489.
- [37] J. H. Sudati et al., "Valeriana officinalis attenuates the rotenone-induced toxicity in drosophila melanogaster," *Neurotoxicology*, vol. 37, pp. 118 – 126, 2013, doi: 10.1016/j.neuro.2013.04.006.
- [38] A. Y. Chen, P. Wilburn, X. Hao, and T. Tully, "Walking deficits and centrophobism in an α -synuclein fly model of Parkinson's disease," *Genes, Brain Behav.*, vol. 13, no. 8, pp. 812 – 820, 2014, doi: 10.1111/gbb.12172.
- [39] L. F. Belfiori-Carrasco, M. S. Marcora, N. I. Bocai, M. F. Ceriani, L. Morelli, and E. M. Castaño, "A novel genetic screen identifies modifiers of age-dependent amyloid β toxicity in the *Drosophila* brain," *Front. Aging Neurosci.*, vol. 9, no. MAR, 2017, doi: 10.3389/fnagi.2017.00061.
- [40] I. Rogers, F. Kerr, P. Martinez, J. Hardy, S. Lovestone, and L. Partridge, "Ageing increases vulnerability to A β 42 toxicity in *Drosophila*," *PLoS One*, vol. 7, no. 7, 2012, doi: 10.1371/journal.pone.0040569.
- [41] S. Burnouf, M. K. Gorsky, J. Dols, S. Grönke, and L. Partridge, "A β 43 is neurotoxic and primes aggregation of A β 40 in vivo," *Acta Neuropathol.*, vol. 130, no. 1, pp. 35 – 47, 2015, doi: 10.1007/s00401-015-1419-y.
- [42] R. Xue et al., "dNAGLU extends life span and promotes fitness and stress resistance in *Drosophila*," *Int. J. Mol. Sci.*, vol. 23, no. 22, 2022, doi: 10.3390/ijms232214433.
- [43] R. Mattioli et al., "Anti-inflammatory activity of a polyphenolic extract from arabidopsis thaliana in in vitro and in vivo models of alzheimer's disease," *Int. J. Mol. Sci.*, vol. 20, no. 3, 2019, doi: 10.3390/ijms20030708.
- [44] J. Tower et al., "Behavioral and molecular markers of death in *Drosophila melanogaster*," *Exp. Gerontol.*, vol. 126, 2019, doi: 10.1016/j.exger.2019.110707.
- [45] Y. H. Siddique, F. Naz, Rahul, and H. Varshney, "Comparative study of rivastigmine and galantamine on the transgenic *Drosophila* model of Alzheimer's disease," *Curr. Res. Pharmacol. Drug Discov.*, vol. 3, 2022, doi: 10.1016/j.crphar.2022.100120.
- [46] S. Burnouf et al., "Deletion of endogenous Tau proteins is not detrimental in *Drosophila*," *Sci. Rep.*, vol. 6, 2016, doi: 10.1038/srep23102.
- [47] A. Gandini et al., "Discovery of dual A β /Tau inhibitors and evaluation of their therapeutic effect on a *Drosophila* model of Alzheimer's disease," *ACS Chem. Neurosci.*, vol. 13, no. 23, pp. 3314 – 3329, 2022, doi: 10.1021/acscchemneuro.2c00357.
- [48] X. Zhang et al., "Downregulation of RBO-PI4KIII α facilitates A β 42 secretion

- and ameliorates neural deficits in A β 42-expressing *Drosophila*,” *J. Neurosci.*, vol. 37, no. 19, pp. 4928 – 4941, 2017, doi: 10.1523/JNEUROSCI.3567-16.2017.
- [49] G. Wei et al., “Enzyme-assisted solvent extraction of high-yield paeonia suffruticosa andr. Seed oil and fatty acid composition and anti-alzheimer’s disease activity,” *J. Oleo Sci.*, vol. 70, no. 8, pp. 1133 – 1146, 2021, doi: 10.5650/jos.ess21040.
- [50] A. Maggiore et al., “Neuroprotective effects of PARP inhibitors in *Drosophila* models of Alzheimer’s disease,” *Cells*, vol. 11, no. 8, 2022, doi: 10.3390/cells11081284.
- [51] M. Arnés, N. Romero, S. Casas-Tintó, Á. Acebes, and A. Ferrús, “PI3K activation prevents A β 42-induced synapse loss and favors insoluble amyloid deposit formation,” *Mol. Biol. Cell*, vol. 31, no. 4, pp. 244 – 260, 2020, doi: 10.1091/mbc.E19-05-0303.
- [52] D. M. Long et al., “Relationships between the circadian system and Alzheimer’s disease-like symptoms in *Drosophila*,” *PLoS One*, vol. 9, no. 8, 2014, doi: 10.1371/journal.pone.0106068.
- [53] J. Thomas et al., “The angiotensin-converting enzyme inhibitor lisinopril mitigates memory and motor deficits in a *drosophila* model of alzheimer’s disease,” *Pathophysiology*, vol. 28, no. 2, pp. 307 – 319, 2021, doi: 10.3390/pathophysiology28020020.
- [54] S. Lee et al., “The calcineurin inhibitor Sarah (Nebula) exacerbates A β 42 phenotypes in a *Drosophila* model of Alzheimer’s disease,” *DMM Dis. Model. Mech.*, vol. 9, no. 3, pp. 295 – 306, 2016, doi: 10.1242/dmm.018069.
- [55] C. De Gregorio, R. Delgado, A. Ibacache, J. Sierralta, and A. Couve, “*Drosophila* Atlastin in motor neurons is required for locomotion and presynaptic function,” *J. Cell Sci.*, vol. 130, no. 20, pp. 3507–3516, Oct. 2017, doi: 10.1242/jcs.201657.
- [56] V. Billea et al., “AUTEN-67 (autophagy enhancer-67) hampers the progression of neurodegenerative symptoms in a *Drosophila* model of Huntington’s disease,” *J. Huntingtons. Dis.*, vol. 5, no. 2, pp. 133 – 147, 2016, doi: 10.3233/JHD-150180.
- [57] J. Varga, N. P. Dér, N. Zsindely, and L. Bodai, “Green tea infusion alleviates neurodegeneration induced by mutant Huntingtin in *Drosophila*,” *Nutr. Neurosci.*, vol. 23, no. 3, pp. 183 – 189, 2020, doi: 10.1080/1028415X.2018.1484021.
- [58] S. J. Banerjee et al., “iPLA2-VIA is required for healthy aging of neurons, muscle, and the female germline in *Drosophila melanogaster*,” *PLoS One*, vol. 16, no. 9 September, 2021, doi: 10.1371/journal.pone.0256738.
- [59] H. H. Abdulbaki and M. A. Al-Deeb, “Chlorpyrifos-induced dopaminergic damage in *Drosophila melanogaster* assessed by gene expression, AChE assay, and negative geotaxis using a new feeding device,” *Genet. Mol. Res.*, vol. 21, no. 3, 2022, doi: 10.4238/gmr19056.
- [60] J.-C. Chang and D. B. Morton, “*Drosophila* lines with mutant and wild type human TDP-43 replacing the endogenous gene reveals phosphorylation and ubiquitination in mutant lines in the absence of viability or lifespan defects,” *PLoS One*, vol. 12, no. 7, 2017, doi: 10.1371/journal.pone.0180828.
- [61] R. T. Yost et al., “Abnormal social interactions in a *Drosophila* mutant of an autism candidate gene: Neuroligin 3,” *Int. J. Mol. Sci.*, vol. 21, no. 13, pp. 1 – 20, 2020, doi: 10.3390/ijms21134601.
- [62] D. Damschroder, C. Reynolds, and R. Wessells, “*Drosophila* tafazzin mutants have impaired exercise capacity,” *Physiol. Rep.*, vol. 6, no. 3, 2018, doi: 10.14814/phy2.13604.
- [63] H. Sasayama et al., “Knockdown of the *Drosophila* fused in sarcoma (FUS)

- homologue causes deficient locomotive behavior and shortening of motoneuron terminal branches,” *PLoS One*, vol. 7, no. 6, 2012, doi: 10.1371/journal.pone.0039483.
- [64] A. M. Thackray et al., “Clearance of variant Creutzfeldt-Jakob disease prions in vivo by the Hsp70 disaggregase system,” *Brain*, vol. 145, no. 9, pp. 3236 – 3249, 2022, doi: 10.1093/brain/awac144.
- [65] W. Gu et al., “Regular exercise rescues heart function defects and shortens the lifespan of *Drosophila* caused by dMnM downregulation,” *Int. J. Environ. Res. Public Health*, vol. 19, no. 24, 2022, doi: 10.3390/ijerph192416554.
- [66] E. L. Ryan, B. DuBoff, M. B. Feany, and J. L. Fridovich-Keil, “Mediators of a long-term movement abnormality in a *Drosophila melanogaster* model of classic galactosemia,” *DMM Dis. Model. Mech.*, vol. 5, no. 6, pp. 796 – 803, 2012, doi: 10.1242/dmm.009050.
- [67] J. Deng et al., “Reducing the excess activin signaling rescues muscle degeneration in myotonic dystrophy type 2 *Drosophila* model,” *J. Pers. Med.*, vol. 12, no. 3, 2022, doi: 10.3390/jpm12030385.
- [68] A. Sujkowski, K. Richardson, M. V Prifti, R. J. Wessells, and S. V Todi, “Endurance exercise ameliorates phenotypes in *Drosophila* models of spinocerebellar ataxias,” *Elife*, vol. 11, 2022, doi: 10.7554/ELIFE.75389.
- [69] A. M. Kaynar et al., “Cost of surviving sepsis: a novel model of recovery from sepsis in *Drosophila melanogaster*,” *Intensive Care Med. Exp.*, vol. 4, no. 1, pp. 1 – 16, 2016, doi: 10.1186/s40635-016-0075-4.
- [70] M. H. Al-Sabri et al., “Statins induce locomotion and muscular phenotypes in *Drosophila melanogaster* that are reminiscent of human myopathy: Evidence for the role of the chloride channel inhibition in the muscular phenotypes,” *Cells*, vol. 11, no. 22, 2022, doi: 10.3390/cells11223528.
- [71] N. Baenas and A. E. Wagner, “*Drosophila melanogaster* as a model organism for obesity and type-2 diabetes mellitus by applying high-sugar and high-fat diets,” *Biomolecules*, vol. 12, no. 2, 2022, doi: 10.3390/biom12020307.
- [72] L. Tamberg et al., “Daughterless, the *Drosophila* orthologue of TCF4, is required for associative learning and maintenance of the synaptic proteome,” *DMM Dis. Model. Mech.*, vol. 13, no. 7, 2020, doi: 10.1242/dmm.042747.
- [73] T. C. Moulin, F. Ferro, A. Hoyer, P. Cheung, M. J. Williams, and H. B. Schiöth, “The *Drosophila melanogaster* levodopa-induced depression model exhibits negative geotaxis deficits and differential gene expression in males and females,” *Front. Neurosci.*, vol. 15, 2021, doi: 10.3389/fnins.2021.653470.
- [74] E. R. Reynolds, “Shortened lifespan and other age-related defects in bang sensitive mutants of *Drosophila melanogaster*,” *G3 Genes, Genomes, Genet.*, vol. 8, no. 12, pp. 3953 – 3960, 2018, doi: 10.1534/g3.118.200610.
- [75] W. H. Eschenbacher et al., “Two rare human mitofusin 2 mutations alter mitochondrial dynamics and induce retinal and cardiac pathology in *Drosophila*,” *PLoS One*, vol. 7, no. 9, 2012, doi: 10.1371/journal.pone.0044296.
- [76] M. Zhao et al., “Knockdown of genes involved in axonal transport enhances the toxicity of human neuromuscular disease-linked MATR3 mutations in *Drosophila*,” *FEBS Lett.*, vol. 594, no. 17, pp. 2800 – 2818, 2020, doi: 10.1002/1873-3468.13858.
- [77] A. M. Muir et al., “Bi-allelic loss-of-function variants in NUP188 cause a recognizable syndrome characterized by neurologic, ocular, and cardiac abnormalities,” *Am. J. Hum. Genet.*, vol.

- 106, no. 5, pp. 623 – 631, 2020, doi: 10.1016/j.ajhg.2020.03.009.
- [78] B. Zhang, Z. Ding, L. Li, L.-K. Xie, Y.-J. Fan, and Y.-Z. Xu, “Two oppositely-charged *sf3b1* mutations cause defective development, impaired immune response, and aberrant selection of intronic branch sites in *Drosophila*,” *PLoS Genet.*, vol. 17, no. 11, 2021, doi: 10.1371/journal.pgen.1009861.
- [79] A. Sujkowski, S. Rainier, J. K. Fink, and R. J. Wessells, “Delayed induction of human NTE (PNPLA6) rescues neurodegeneration and mobility defects of *drosophila* swiss cheese (SWS) mutants,” *PLoS One*, vol. 10, no. 12, 2015, doi: 10.1371/journal.pone.0145356.
- [80] M. J. Ferreira et al., “*Drosophila melanogaster* white mutant *w1118* undergo retinal degeneration,” *Front. Neurosci.*, vol. 11, no. JAN, 2018, doi: 10.3389/fnins.2017.00732.
- [81] R. R. R. Duarte et al., “The psychiatric risk gene *nt5c2* regulates adenosine monophosphate-activated protein kinase signaling and protein translation in human neural progenitor cells,” *Biol. Psychiatry*, vol. 86, no. 2, pp. 120 – 130, 2019, doi: 10.1016/j.biopsych.2019.03.977.
- [82] S. Mendez et al., “The TreadWheel: A novel apparatus to measure genetic variation in response to gently induced exercise for *Drosophila*,” *PLoS One*, vol. 11, no. 10, 2016, doi: 10.1371/journal.pone.0164706.
- [83] S. P. E S, R. P. Nayak, P. Saldanha, M. B J, and A. Prabhu, “Neuroprotective activity of pyrazolone derivatives against paraquat-induced oxidative stress and locomotor impairment in *Drosophila melanogaster*,” *Int. J. Curr. Res. Rev.*, vol. 12, no. 23, pp. 68–75, 2020, doi: 10.31782/IJCRR.2020.122329.
- [84] C. Li et al., “Dysfunction of GRAP, encoding the GRB2-related adaptor protein, is linked to sensorineural hearing loss,” *Proc. Natl. Acad. Sci. U. S. A.*, vol. 116, no. 4, pp. 1347 – 1352, 2019, doi: 10.1073/pnas.1810951116.
- [85] E. J. Shah, K. Gurdziel, and D. M. Ruden, “*Drosophila* exhibit divergent sex-based responses in transcription and motor function after traumatic brain injury,” *Front. Neurol.*, vol. 11, 2020, doi: 10.3389/fneur.2020.00511.
- [86] E. J. Shah, K. Gurdziel, and D. M. Ruden, “Sex-differences in traumatic brain injury in the absence of tau in *Drosophila*,” *Genes (Basel)*, vol. 12, no. 6, 2021, doi: 10.3390/genes12060917.
- [87] E. N. Anderson et al., “Traumatic injury compromises nucleocytoplasmic transport and leads to TDP-43 pathology,” *Elife*, vol. 10, 2021, doi: 10.7554/eLife.67587.
- [88] V. Chauhan and A. Chauhan, “Traumatic injury in female *Drosophila melanogaster* affects the development and induces behavioral abnormalities in the offspring,” *Behav. Brain Funct.*, vol. 15, no. 1, 2019, doi: 10.1186/s12993-019-0163-1.
- [89] P. Xu et al., “Atg2, Atg9 and Atg18 in mitochondrial integrity, cardiac function and healthspan in *Drosophila*,” *J. Mol. Cell. Cardiol.*, vol. 127, pp. 116 – 124, 2019, doi: 10.1016/j.yjmcc.2018.12.006.
- [90] J. C. Zheng et al., “Secretory Carrier Membrane Protein (SCAMP) deficiency influences behavior of adult flies,” *Front. Cell Dev. Biol.*, vol. 2, no. NOV, 2014, doi: 10.3389/fcell.2014.00064.
- [91] G. A. Kaas et al., “Lithium-responsive seizure-like hyperexcitability is caused by a mutation in the *drosophila* voltage-gated sodium channel gene *paralytic*,” *eNeuro*, vol. 3, no. 5, 2016, doi: 10.1523/ENEURO.0221-16.2016.
- [92] E. V Ryabova et al., “Morpho-functional consequences of swiss cheese knockdown in glia of *drosophila melanogaster*,” *Cells*, vol. 10, no. 3, pp. 1 – 20, 2021, doi: 10.3390/cells10030529.

- [93] J. Azpurua, E. G. El-Karim, M. Tranquille, and J. Dubnau, "A behavioral screen for mediators of age-dependent TDP-43 neurodegeneration identifies SF2/SRSF1 among a group of potent suppressors in both neurons and glia," *PLoS Genet.*, vol. 17, no. 11, 2021, doi: 10.1371/journal.pgen.1009882.
- [94] E. P. Ratliff et al., "Aging and autophagic function influences the progressive decline of adult *Drosophila* behaviors," *PLoS One*, vol. 10, no. 7, 2015, doi: 10.1371/journal.pone.0132768.
- [95] M. A. Jones, J. W. Gargano, D. Rhodenizer, I. Martin, P. Bhandari, and M. Grotewiel, "A forward genetic screen in *Drosophila* implicates insulin signaling in age-related locomotor impairment," *Exp. Gerontol.*, vol. 44, no. 8, pp. 532 – 540, 2009, doi: 10.1016/j.exger.2009.05.007.
- [96] S. Liao, S. Broughton, and D. R. Nässel, "Behavioral senescence and aging-related changes in motor neurons and brain neuromodulator levels are ameliorated by lifespan-extending reproductive dormancy in *Drosophila*," *Front. Cell. Neurosci.*, vol. 11, 2017, doi: 10.3389/fncel.2017.00111.
- [97] D. Rhodenizer, I. Martin, P. Bhandari, S. D. Pletcher, and M. Grotewiel, "Genetic and environmental factors impact age-related impairment of negative geotaxis in *Drosophila* by altering age-dependent climbing speed," *Exp. Gerontol.*, vol. 43, no. 8, pp. 739 – 748, 2008, doi: 10.1016/j.exger.2008.04.011.
- [98] A. Bednářová, A. Tomčala, M. Mochanová, D. Kodrík, and N. Krishnan, "Disruption of adipokinetic hormone mediated energy homeostasis has subtle effects on physiology, behavior and lipid status during aging in *Drosophila*," *Front. Physiol.*, vol. 9, no. JUL, 2018, doi: 10.3389/fphys.2018.00949.
- [99] A. Ueda et al., "Two novel forms of ERG oscillation in *Drosophila*: age and activity dependence," *J. Neurogenet.*, vol. 32, no. 2, pp. 118 – 126, 2018, doi: 10.1080/01677063.2018.1461866.
- [100] P. Shahrestani, J. Quach, L. D. Mueller, and M. R. Rose, "Paradoxical physiological transitions from aging to late life in *Drosophila*," *Rejuvenation Res.*, vol. 15, no. 1, pp. 49 – 58, 2012, doi: 10.1089/rej.2011.1201.
- [101] R. F. Chan et al., "Contrasting influences of *Drosophila* white/mini-white on ethanol sensitivity in two different behavioral assays," *Alcohol. Clin. Exp. Res.*, vol. 38, no. 6, pp. 1582 – 1593, 2014, doi: 10.1111/acer.12421.
- [102] A. Raj, P. Shah, and N. Agrawal, "Sedentary behavior and altered metabolic activity by AgNPs ingestion in *Drosophila melanogaster*," *Sci. Rep.*, vol. 7, no. 1, 2017, doi: 10.1038/s41598-017-15645-6.
- [103] S.-H. Lee, H.-Y. Lee, E.-J. Lee, D. Khang, and K.-J. Min, "Effects of carbon nanofiber on physiology of *Drosophila*," *Int. J. Nanomedicine*, vol. 10, pp. 3687 – 3697, 2015, doi: 10.2147/IJN.S82637.
- [104] P. K. Mishra et al., "Wood-based cellulose nanofibrils: Haemocompatibility and impact on the development and behaviour of *Drosophila melanogaster*," *Biomolecules*, vol. 9, no. 8, 2019, doi: 10.3390/biom9080363.
- [105] S. Yan et al., "Chronic exposure to the star polycation (SPc) nanocarrier in the larval stage adversely impairs life history traits in *Drosophila melanogaster*," *J. Nanobiotechnology*, vol. 20, no. 1, 2022, doi: 10.1186/s12951-022-01705-1.
- [106] K. Sood, J. Kaur, H. Singh, S. Kumar Arya, and M. Khatri, "Comparative toxicity evaluation of graphene oxide (GO) and zinc oxide (ZnO) nanoparticles on *Drosophila melanogaster*," *Toxicol. Reports*, vol. 6, pp. 768 – 781, 2019, doi: 10.1016/j.toxrep.2019.07.009.
- [107] N. Songvorawit, P. Phengphuang, and T.

- Khongkhieo, “Fluorescent silica nanoparticles as an internal marker in fruit flies and their effects on survivorship and fertility,” *Sci. Rep.*, vol. 12, no. 1, 2022, doi: 10.1038/s41598-022-24301-7.
- [108] J. L. Podratz et al., “An automated climbing apparatus to measure chemotherapy-induced neurotoxicity in *Drosophila melanogaster*,” *Fly (Austin)*, vol. 7, no. 3, pp. 187 – 192, 2013, doi: 10.4161/fly.24789.
- [109] J. L. Podratz et al., “*Drosophila melanogaster*: A new model to study cisplatin-induced neurotoxicity,” *Neurobiol. Dis.*, vol. 43, no. 2, pp. 330 – 337, 2011, doi: 10.1016/j.nbd.2011.03.022.
- [110] B. M. Baker et al., “The *Drosophila* brain on cocaine at single-cell resolution,” *Genome Res.*, vol. 31, no. 10, pp. 1927 – 1937, 2021, doi: 10.1101/gr.268037.120.
- [111] A. K. Lima, H. Dhillon, and A. R. Dillman, “ShK-domain-containing protein from a parasitic nematode modulates *Drosophila melanogaster* immunity,” *Pathogens*, vol. 11, no. 10, 2022, doi: 10.3390/pathogens11101094.
- [112] M. R. Poetini et al., “Iron overload during the embryonic period develops hyperactive like behavior and dysregulation of biogenic amines in *Drosophila melanogaster*,” *Dev. Biol.*, vol. 475, pp. 80 – 90, 2021, doi: 10.1016/j.ydbio.2021.03.006.
- [113] M. P. Singh et al., “Protection of Phytoextracts against Rotenone-Induced Organismal Toxicities in *Drosophila melanogaster* via the Attenuation of ROS Generation,” *Appl. Sci.*, vol. 12, no. 19, 2022, doi: 10.3390/app12199822.
- [114] Z. Chen, F. Wang, D. Wen, and R. Mu, “Exposure to bisphenol A induced oxidative stress, cell death and impaired epithelial homeostasis in the adult *Drosophila melanogaster* midgut,” *Ecotoxicol. Environ. Saf.*, vol. 248, 2022, doi: 10.1016/j.ecoenv.2022.114285.
- [115] F. H. Figueira et al., “Exposure to atrazine alters behaviour and disrupts the dopaminergic system in *Drosophila melanogaster*,” *Comp. Biochem. Physiol. Part - C Toxicol. Pharmacol.*, vol. 202, pp. 94 – 102, 2017, doi: 10.1016/j.cbpc.2017.08.005.
- [116] P. C. Lovejoy and A. C. Fiumera, “Effects of dual exposure to the herbicides atrazine and paraquat on adult climbing ability and longevity in *Drosophila melanogaster*,” *Insects*, vol. 10, no. 11, 2019, doi: 10.3390/insects10110398.
- [117] X. Qiao et al., “An insecticide target in mechanoreceptor neurons,” *Sci. Adv.*, vol. 8, no. 47, 2022, doi: 10.1126/sciadv.abq3132.
- [118] T. D. Algarve, C. E. Assmann, T. Aigaki, and I. B. M. da Cruz, “Parental and preimaginal exposure to methylmercury disrupts locomotor activity and circadian rhythm of adult *Drosophila melanogaster*,” *Drug Chem. Toxicol.*, vol. 43, no. 3, pp. 255–265, 2020, doi: 10.1080/01480545.2018.1485689.
- [119] T. O. Johnson et al., “Benzo[a]pyrene and Benzo[a]pyrene-7,8-dihydrodiol-9,10-epoxide induced locomotor and reproductive senescence and altered biochemical parameters of oxidative damage in Canton-S *Drosophila melanogaster*,” *Toxicol. Reports*, vol. 8, pp. 571 – 580, 2021, doi: 10.1016/j.toxrep.2021.03.001.
- [120] P. Subramanian, K. Kaliyamoorthy, J. J. Jayapalan, P. Shafinaz Abdul-Rahman, and O. H. Hashim, “Influence of quercetin in the temporal regulation of redox homeostasis in *drosophila melanogaster*,” *J. Insect Sci.*, vol. 17, no. 2, 2017, doi: 10.1093/jisesa/iex040.
- [121] F. Valéria Soares de Araújo Pinho et al., “Phytochemical constituents and toxicity of *duguetia furfuracea* hydroalcoholic extract in *Drosophila melanogaster*,” *Evid. Based. Complement. Alternat. Med.*, vol. 2014, p. 838101, 2014, doi: 10.1155/2014/838101.

- [122] B. Fernández-Puntero, I. Barroso, I. Iglesias, J. Benedí, and A. Villar, "Antioxidant activity of Fraxetin: In vivo and ex vivo parameters in normal situation versus induced stress," *Biol. Pharm. Bull.*, vol. 24, no. 7, pp. 777 – 784, 2001, doi: 10.1248/bpb.24.777.
- [123] G. J. T. Salazar, A. Ecker, S. A. Adefegha, and J. G. M. da Costa, "Advances in evaluation of antioxidant and toxicological properties of *stryphnodendron rotundifolium* mart. in *Drosophila melanogaster* model," *Foods*, vol. 11, no. 15, 2022, doi: 10.3390/foods11152236.
- [124] S. P. Balasubramani et al., "Pomegranate juice enhances healthy lifespan in *Drosophila melanogaster*: An exploratory study," *Front. Public Heal.*, vol. 2, no. DEC, 2014, doi: 10.3389/fpubh.2014.00245.
- [125] Q. Wang et al., "Soft-shelled turtle peptides extend lifespan and healthspan in *Drosophila*," *Nutrients*, vol. 14, no. 24, 2022, doi: 10.3390/nu14245205.
- [126] S. N. Prasad and Muralidhara, "Neuromodulatory effects of aqueous extract of *Coriandrum sativum* seeds against acrylamide induced toxicity in *Drosophila melanogaster*," *Int. J. Res. Pharm. Sci.*, vol. 10, no. 2, pp. 1127 – 1135, 2019, doi: 10.26452/ijrps.v10i2.393.
- [127] P. Bhandari, M. A. Jones, I. Martin, and M. S. Grotewiel, "Dietary restriction alters demographic but not behavioral aging in *Drosophila*," *Aging Cell*, vol. 6, no. 5, pp. 631 – 637, 2007, doi: 10.1111/j.1474-9726.2007.00320.x.
- [128] K. A. Wilson et al., "GWAS for lifespan and decline in climbing ability in flies upon dietary restriction reveal decima as a mediator of insulin-like peptide production," *Curr. Biol.*, vol. 30, no. 14, pp. 2749 – 2760.e3, 2020, doi: 10.1016/j.cub.2020.05.020.
- [129] D. C. Lee et al., "Dietary supplementation with the ketogenic diet metabolite beta-hydroxybutyrate ameliorates post-tbi aggression in young-adult male *drosophila*," *Front. Neurosci.*, vol. 13, no. OCT, 2019, doi: 10.3389/fnins.2019.01140.
- [130] J. Ro et al., "Serotonin signaling mediates protein valuation and aging," *Elife*, vol. 5, no. AUGUST, 2016, doi: 10.7554/eLife.16843.
- [131] Y. Su et al., "Alpha-ketoglutarate extends *Drosophila* lifespan by inhibiting mTOR and activating AMPK," *Aging (Albany. NY)*, vol. 11, no. 12, pp. 4183 – 4197, 2019, doi: 10.18632/aging.102045.
- [132] J. Jung, D.-I. Kim, G.-Y. Han, and H. W. Kwon, "The effects of high fat diet-induced stress on olfactory sensitivity, behaviors, and transcriptional profiling in *Drosophila melanogaster*," *Int. J. Mol. Sci.*, vol. 19, no. 10, 2018, doi: 10.3390/ijms19102855.
- [133] R. P. J. Cormier, C. M. Champigny, C. J. Simard, P.-D. St-Coeur, and N. Pichaud, "Dynamic mitochondrial responses to a high-fat diet in *Drosophila melanogaster*," *Sci. Rep.*, vol. 9, no. 1, 2019, doi: 10.1038/s41598-018-36060-5.
- [134] S. Liao, M. Amcoff, and D. R. Nässel, "Impact of high-fat diet on lifespan, metabolism, fecundity and behavioral senescence in *Drosophila*," *Insect Biochem. Mol. Biol.*, vol. 133, 2021, doi: 10.1016/j.ibmb.2020.103495.
- [135] N. Wongchum, A. Dechakhamphu, P. Panya, S. Pinlaor, S. Pinmongkhonkul, and A. Tanomtong, "Hydroethanolic *Cyperus rotundus* L. extract exhibits anti-obesity property and increases lifespan expectancy in *Drosophila melanogaster* fed a high-fat diet," *J. HerbMed Pharmacol.*, vol. 11, no. 2, pp. 296 – 304, 2022, doi: 10.34172/jhp.2022.35.
- [136] O. Rivera, L. McHan, B. Konadu, S. Patel, S. Sint Jago, and M. E. Talbert, "A high-fat diet impacts memory and gene expression of the head in mated female

- Drosophila melanogaster*,” *J. Comp. Physiol. B Biochem. Syst. Environ. Physiol.*, vol. 189, no. 2, pp. 179 – 198, 2019, doi: 10.1007/s00360-019-01209-9.
- [137] D.-T. Wen, W.-Q. Wang, W.-Q. Hou, S.-X. Cai, and S.-S. Zhai, “Endurance exercise protects aging *Drosophila* from high-salt diet (HSD)-induced climbing capacity decline and lifespan decrease by enhancing antioxidant capacity,” *Biol. Open*, vol. 9, no. 5, 2020, doi: 10.1242/bio.045260.
- [138] K. I. Kasozi et al., “Low concentrations of monosodium glutamate (MSG) are safe in male *Drosophila melanogaster*,” *BMC Res. Notes*, vol. 11, no. 1, 2018, doi: 10.1186/s13104-018-3775-x.
- [139] I. Hubrecht, N. Baenas, C. Sina, and A. E. Wagner, “Effects of non-caloric artificial sweeteners on naïve and dextran sodium sulfate-exposed *Drosophila melanogaster*,” *Food Front.*, vol. 3, no. 4, pp. 728 – 735, 2022, doi: 10.1002/fft2.147.
- [140] E. Gómez et al., “Impact of probiotics on development and behaviour in *Drosophila melanogaster* -a potential in vivo model to assess probiotics,” *Benef. Microbes*, vol. 10, no. 2, pp. 179 – 188, 2019, doi: 10.3920/BM2018.0012.
- [141] A. Sujkowski, B. Bazzell, K. Carpenter, R. Arking, and R. J. Wessells, “Endurance exercise and selective breeding for longevity extend *Drosophila* healthspan by overlapping mechanisms,” *Aging (Albany. NY.)*, vol. 7, no. 8, pp. 535 – 552, 2015, doi: 10.18632/aging.100789.
- [142] L. Zheng, Y. Feng, D. T. Wen, H. Wang, and X. S. Wu, “Fatiguing exercise initiated later in life reduces incidence of fibrillation and improves sleep quality in *Drosophila*,” *Age (Omaha.)*, vol. 37, no. 4, 2015, doi: 10.1007/s11357-015-9816-7.
- [143] N. Piazza, B. Gosangi, S. Devilla, R. Arking, and R. Wessells, “Exercise-training in young *Drosophila melanogaster* reduces age-related decline in mobility and cardiac performance,” *PLoS One*, vol. 4, no. 6, 2009, doi: 10.1371/journal.pone.0005886.
- [144] M. Ding, L. Zheng, Q. F. Li, W. L. Wang, W. Da Peng, and M. Zhou, “Exercise-training regulates apolipoprotein B in *Drosophila* to improve HFD-mediated cardiac function damage and low exercise capacity,” *Front. Physiol.*, vol. 12, 2021, doi: 10.3389/fphys.2021.650959.
- [145] G. Fedele, E. W. Green, E. Rosato, and C. P. Kyriacou, “An electromagnetic field disrupts negative geotaxis in *Drosophila* via a CRY-dependent pathway,” *Nat. Commun.*, vol. 5, 2014, doi: 10.1038/ncomms5391.
- [146] J.-E. Bae et al., “Positive geotactic behaviors induced by geomagnetic field in *Drosophila*,” *Mol. Brain*, vol. 9, no. 1, 2016, doi: 10.1186/s13041-016-0235-1.
- [147] R. J. Schilder and M. Raynor, “Molecular plasticity and functional enhancements of leg muscles in response to hypergravity in the fruit fly *Drosophila melanogaster*,” *J. Exp. Biol.*, vol. 220, no. 19, pp. 3508 – 3518, 2017, doi: 10.1242/jeb.160523.
- [148] C. S. Kim et al., “Chronic low-dose γ -irradiation of *Drosophila melanogaster* larvae induces gene expression changes and enhances locomotive behavior,” *J. Radiat. Res.*, vol. 56, no. 3, pp. 475 – 484, 2015, doi: 10.1093/jrr/rru128.
- [149] L. J. Sudmeier, S. P. Howard, and B. Ganetzky, “A *Drosophila* model to investigate the neurotoxic side effects of radiation exposure,” *DMM Dis. Model. Mech.*, vol. 8, no. 7, pp. 669 – 677, 2015, doi: 10.1242/dmm.019786.
- [150] Y. Xia, W. Xu, S. Meng, N. K. H. Lim, W. Wang, and F.-D. Huang, “An efficient and reliable assay for investigating the effects of hypoxia/anoxia on *Drosophila*,” *Neurosci. Bull.*, vol. 34, no. 2, pp. 397 – 402, 2018, doi: 10.1007/s12264-017-0173-7.
- [151] A. Bretman and C. Fricke, “Exposure to

- males, but not receipt of sex peptide, accelerates functional ageing in female fruit flies,” *Funct. Ecol.*, vol. 33, no. 8, pp. 1459 – 1468, 2019, doi: 10.1111/1365-2435.13339.
- [152] I. Krams et al., “Short-term exposure to predation affects body elemental composition, climbing speed and survival ability in *Drosophila melanogaster*,” *PeerJ*, vol. 2016, no. 4, 2016, doi: 10.7717/PEERJ.2314.
- [153] A. R. McNeil et al., “Conditions affecting social space in *Drosophila melanogaster*,” *J. Vis. Exp.*, vol. 2015, no. 105, 2015, doi: 10.3791/53242.
- [154] I. Schoberleitner, “Regulation of sensory perception and motor abilities by brain-specific action of chromatin remodeling factor CHD1,” *Front. Mol. Neurosci.*, vol. 15, 2022, doi: 10.3389/fnmol.2022.840966.
- [155] Y. Sun, L. Liu, Y. Ben-Shahar, J. S. Jacobs, D. F. Eberl, and M. J. Welsh, “TRPA channels distinguish gravity sensing from hearing in Johnston’s organ,” *Proc. Natl. Acad. Sci. U. S. A.*, vol. 106, no. 32, pp. 13606 – 13611, 2009, doi: 10.1073/pnas.0906377106.
- [156] P. G. M’Angale and B. E. Staveley, “Knockdown of the putative Lifeguard homologue CG3814 in neurons of *Drosophila melanogaster*,” *Genet. Mol. Res.*, vol. 15, no. 4, 2016, doi: 10.4238/gmr15049290.
- [157] Z. Zhang et al., “METTL14 Regulates Intestine Cellular Senescence through m6A Modification of Lamin B Receptor,” *Oxid. Med. Cell. Longev.*, vol. 2022, 2022, doi: 10.1155/2022/9096436.
- [158] J. Fernandes and J. Varghese, “Sexually dimorphic microRNA miR-190 regulates lifespan in male *Drosophila*,” *RNA Biol.*, vol. 19, no. 1, pp. 1085 – 1093, 2022, doi: 10.1080/15476286.2022.2127544.
- [159] J. Sun et al., “Neural control of startle-induced locomotion by the mushroom bodies and associated neurons in *drosophila*,” *Front. Syst. Neurosci.*, vol. 12, 2018, doi: 10.3389/fnsys.2018.00006.
- [160] A. Eriksson et al., “Neuromodulatory circuit effects on *Drosophila* feeding behaviour and metabolism,” *Sci. Rep.*, vol. 7, no. 1, 2017, doi: 10.1038/s41598-017-08466-0.
- [161] H. Augustin, J. Adcott, C. J. H. Elliott, and L. Partridge, “Complex roles of myoglianin in regulating adult performance and lifespan,” *Fly (Austin)*, vol. 11, no. 4, pp. 284 – 289, 2017, doi: 10.1080/19336934.2017.1369638.
- [162] I. Devambez et al., “Identification of DmTLL5 as a major tubulin glutamylase in the *Drosophila* nervous system,” *Sci. Rep.*, vol. 7, no. 1, 2017, doi: 10.1038/s41598-017-16586-w.
- [163] S. E. El-Kholy, B. Affi, I. El-Husseiny, and A. Seif, “Octopamine signaling via OctaR is essential for a well-orchestrated climbing performance of adult *Drosophila melanogaster*,” *Sci. Rep.*, vol. 12, no. 1, 2022, doi: 10.1038/s41598-022-18203-x.
- [164] D. L. Clayton, “Circadian and geotactic behaviors: Genetic pleiotropy in *Drosophila melanogaster*,” *J. Circadian Rhythms*, vol. 14, no. 1, pp. 1 – 5, 2016, doi: 10.5334/jcr.140.
- [165] E. S. Pak and A. K. Murashov, “*Drosophila* passive avoidance behavior as a new paradigm to study associative aversive learning,” *J. Vis. Exp.*, vol. 2021, no. 176, 2021, doi: 10.3791/63163.
- [166] N. Dravec et al., “Reduced insulin signaling targeted to serotonergic neurons but not other neuronal subtypes extends lifespan in *Drosophila melanogaster*,” *Front. Aging Neurosci.*, vol. 14, 2022, doi: 10.3389/fnagi.2022.893444.
- [167] M. Z. B. H. Ismail, M. D. Hodges, M. Boylan, R. Achall, A. Shirras, and S. J. Broughton, “The *Drosophila* insulin receptor independently modulates lifespan and locomotor senescence,” *PLoS One*, vol. 10, no. 5, 2015, doi:

- 10.1371/journal.pone.0125312.
- [168] J. Zhu, J.-G. Lee, J. van de Leemput, H. Lee, and Z. Han, “Functional analysis of SARS-CoV-2 proteins in *Drosophila* identifies Orf6-induced pathogenic effects with Selinexor as an effective treatment,” *Cell Biosci.*, vol. 11, no. 1, 2021, doi: 10.1186/s13578-021-00567-8.
- [169] A. Pooryasin and A. Fiala, “Identified serotonin-releasing neurons induce behavioral quiescence and suppress mating in *Drosophila*,” *J. Neurosci.*, vol. 35, no. 37, pp. 12792 – 12812, 2015, doi: 10.1523/JNEUROSCI.1638-15.2015.
- [170] V. L. Barnes, A. Bhat, A. Unnikrishnan, A. R. Heydari, R. Arking, and L. A. Pile, “SIN3 is critical for stress resistance and modulates adult lifespan,” *Aging (Albany. NY)*, vol. 6, no. 8, pp. 645 – 660, 2014, doi: 10.18632/aging.100684.
- [171] D. B. Jørgensen, M. Ørsted, and T. N. Kristensen, “Sustained positive consequences of genetic rescue of fitness and behavioural traits in inbred populations of *Drosophila melanogaster*,” *J. Evol. Biol.*, vol. 35, no. 6, pp. 868 – 878, 2022, doi: 10.1111/jeb.14015.
- [172] E. Bier, “*Drosophila*, the golden bug, emerges as a tool for human genetics,” *Nat. Rev. Genet.*, vol. 6, no. 1, pp. 9–23, Jan. 2005, doi: 10.1038/nrg1503.
- [173] M. E. Fortini, M. P. Skupski, M. S. Boguski, and I. K. Hariharan, “A Survey of human disease gene counterparts in the *Drosophila* genome,” *J. Cell Biol.*, vol. 150, no. 2, pp. F23–F30, Jul. 2000, doi: 10.1083/jcb.150.2.F23.
- [174] B. Ugur, K. Chen, and H. J. Bellen, “*Drosophila* tools and assays for the study of human diseases,” *Dis. Model. Mech.*, vol. 9, no. 3, pp. 235–244, Mar. 2016, doi: 10.1242/dmm.023762.
- [175] S. Yamamoto et al., “A *Drosophila* genetic resource of mutants to study mechanisms underlying human genetic diseases,” *Cell*, vol. 159, no. 1, pp. 200–214, Sep. 2014, doi: 10.1016/j.cell.2014.09.002.
- [176] K. G. Hales, C. A. Korey, A. M. Larracuenta, and D. M. Roberts, “Genetics on the Fly: A primer on the *Drosophila* model system,” *Genetics*, vol. 201, no. 3, pp. 815–842, Nov. 2015, doi: 10.1534/genetics.115.183392.
- [177] C. D. Nichols, J. Becnel, and U. B. Pandey, “Methods to assay *Drosophila* behavior,” *J. Vis. Exp.*, no. 61, Mar. 2012, doi: 10.3791/3795.
- [178] M. D. Rand, “Drosophotoxicology: The growing potential for *Drosophila* in neurotoxicology,” *Neurotoxicol. Teratol.*, vol. 32, no. 1, pp. 74–83, Jan. 2010, doi: 10.1016/j.ntt.2009.06.004