

El-Hayah

JURNAL BIOLOGI

Journal Homepage: <http://ejournal.uin-malang.ac.id/index.php/bio/index>

e-ISSN: 2460-7207, p-ISSN: 2086-0064

Original research article

Adaptive Responses and Differences in Morphophysiological Resistance of *Dendrobium* sp., *Phalaenopsis* sp., and *Vanda* sp. Orchids to Abiotic Stress

Moch. Faizul Huda¹, Ospa Pea Yuanita Meishanti¹, Rossanita Truelovin Hadi Putri¹, Ahmad Sahal Mahfudz¹

¹Biology Education Study Program, Faculty of Education, Universitas KH. A. Wahab Hasbullah, Garuda Street 9, Tambakberas Jombang, Indonesia, 61419

*Corresponding author

Email: mochfaizulhuda@unwaha.ac.id

DOI: [10.18860/elha.v10i4.37654](https://doi.org/10.18860/elha.v10i4.37654)

Article Info

Article history:

Received 21 November 2025

Received in revised form 27

February 2026

Accepted 13 March 2026

Keywords:

Abiotic stress;

Morphophysiological response;

Orchids;

Plant resistance;

Salinity (NaCl)

Abstract

Orchids are an important horticultural commodity in Indonesia with high economic and conservation value; however, their sustainability is threatened by abiotic stress, such as salinity. This study aims to evaluate the resistance of *Dendrobium* sp., *Phalaenopsis* sp., and *Vanda* sp. to NaCl-induced salinity stress using a completely randomized design (CRD) with three replications. The treatments (0%, 1%, 5%, and 10% NaCl) were applied every three days for five weeks under greenhouse conditions. Morphological responses were observed visually, while chlorophyll content (mg/L) was measured spectrophotometrically at 649 and 665 nm. The results showed that chlorophyll content decreased significantly with increasing NaCl concentration. Total chlorophyll of *Phalaenopsis* sp. decreased sharply with increasing NaCl concentration from 5.25 mg/L (control), 3.78 mg/L (1%), 1.74 mg/L (5%) and 0.54 mg/L (10%); In contrast, *Dendrobium* sp. showed fluctuations from 5.02 mg/L (control), 8.20 mg/L (1%), 3.74 mg/L (5%) and 4.05 mg/L (10%). While *Vanda* sp. experienced a moderate decrease from 9.42 mg/L (control), 9.41 mg/L (1%), 4.53 mg/L (5%), and 4.26 mg/L (10%). *Vanda* sp. showed the highest salinity tolerance, followed by *Dendrobium* sp., while *Phalaenopsis* sp. was the most susceptible. NaCl stress significantly reduced chlorophyll content in all orchids.

1. INTRODUCTION

Orchids are a horticultural commodity with high economic value [1]. Market demand for orchids continues to increase year after year [2]. This situation has driven massive exploitation of orchids and their rapid propagation through intensive cultivation [3]. However, in practice, orchid cultivation often faces challenges including drought [4], salinity

[5], and changes in temperature and climate [6]. These factors can inhibit plant growth and development, resulting in serious economic losses.

Abiotic stress has been shown to affect plant growth and the morphological, physiological, and genomic characteristics of orchids both *in vitro* and *in vivo* [3; 7; 8; 9]. Among abiotic stressors, salinity has emerged

as a major constraint on plant productivity worldwide. Salinity stress, primarily caused by the accumulation of sodium chloride (NaCl), disrupts plant physiological and metabolic processes through osmotic stress, ion toxicity, and oxidative damage [10]. These effects lead to reduced water uptake, impaired photosynthesis, including photosynthetic pigment levels, particularly chlorophyll, and ultimately decreased plant growth and yield [11], [12], [13]. Therefore, resistance to abiotic stress (salinity stressors) is key to developing superior orchid varieties [14].

In addition to physiological factors, exposure to abiotic stress affects plant morphology. Morphological responses to abiotic stress are associated with the accumulation of phenolic compounds, which act as secondary metabolites involved in plant defence mechanisms. These compounds play a role in mitigating oxidative stress and maintaining cellular integrity, thereby influencing visible morphological changes such as leaf chlorosis, necrosis, and growth inhibition [9]. The ability to produce secondary metabolites varies among species [15] and can serve as a natural plant defense [16].

Identifying morphological and physiological responses to abiotic stress is crucial as a basis for developing conservation and plant breeding strategies. Studies on the effects of stress on orchid varieties are still very limited [17], particularly the comparison of morphological and physiological responses of three native Indonesian orchid varieties to abiotic stress. The selection of *Dendrobium*, *Phalaenopsis*, and *Vanda* in this study is based on their ecological diversity, physiological variation, and economic importance. Collectively, these three genera represent a gradient of adaptive capacity from tolerant, and intermediate to sensitive. This variation provides a strong scientific basis for comparative analysis aimed at identifying key traits associated with salinity tolerance. This study aimed to determine the morphological and physiological responses of *Dendrobium*

sp., *Phalaenopsis* sp., and *Vanda* sp. orchid varieties to NaCl abiotic stress and to determine the orchid varieties with the best resistance characteristics to NaCl abiotic stress.

2. MATERIALS AND METHODS

This study used an experimental approach with a completely randomized design (CRD) to test orchid resistance to abiotic stress. The study was conducted in a greenhouse and biology laboratory. Samples included *Dendrobium* sp., *Phalaenopsis* sp., and *Vanda* sp., with the independent variable being NaCl abiotic stress and the dependent variable being the plant's resistance level based on morphological and physiological data.

Abiotic Stress Injection

The abiotic stress treatment consisted of injections of NaCl solutions (0%, 1%, 5%, and 10%) every three days for five weeks using a root wounding method using a syringe. A fixed volume of NaCl solution (1 mL per plant) was injected into the root zone to ensure uniform exposure. The root-wounding injection method was selected to facilitate direct uptake of the saline solution and to simulate localized salinity stress conditions, thereby enhancing the consistency and effectiveness of stress induction.

Morphological Data Collection and Chlorophyll Testing

Research documentation was conducted to obtain morphological data and compare the NaCl injection treatment with the control. The chlorophyll test was conducted by grinding 0.5 g of leaves with 10 ml of 96% ethanol until the leaves were pale and the solvent green. The extract solution was then filtered through filter paper and placed in a dark vial. The solution was transferred to a cuvette up to the mark and then read using the absorbance at 649 nm and 665 nm. The chlorophyll a and b levels, as well as total chlorophyll (mg/L) were calculated using the following formula [18]:

$$\text{Chl a (mg/L)} = (13.7 \times A_{665}) - (5.76 \times A_{649})$$

$$\text{Chl b (mg/L)} = (25.80 \times A_{649}) - (7.60 \times A_{665})$$

$$\text{Total chlorophyll (Chl a + Chl b) (mg/L)} = (6.10 \times A_{665} + 20.04 \times A_{649})$$

Data Analysis

The measurement data were analyzed quantitatively using Microsoft Excel software, including the calculation of mean values and standard deviation to evaluate data variability, and were subsequently presented in tables and graphical forms. The standard deviation presented reflects variability among treatment levels.

3. RESULTS

The responses of *Dendrobium* sp., *Phalaenopsis* sp., and *Vanda* sp. to NaCl salinity stress varied significantly, both in terms of chlorophyll content and morphological changes (Table 1; Figure 1). The total chlorophyll content of *Phalaenopsis* sp. decreased drastically with increasing NaCl

concentration. In the control, the chlorophyll content was 5.25 mg/L, which decreased progressively to 3.78 mg/L (1% NaCl), 1.74 mg/L (5% NaCl), and 0.54 mg/L (10% NaCl). This reduction in chlorophyll content was accompanied by observable morphological changes, including leaf chlorosis, necrotic lesions along the leaf margins, and root browning.

A slightly different trend was observed for *Dendrobium* sp. Chlorophyll content increased at 1% NaCl (8.20 mg/L) compared to the control (5.02 mg/L), indicating a stimulatory effect at low salinity levels. However, at higher salinity levels, chlorophyll decreased to 3.74 mg/L (5% NaCl) and 4.05 mg/L (10% NaCl). Morphological observations revealed only mild necrotic spots on the leaves and reduced shoot elongation. These results indicate that *Dendrobium* sp. has a moderate level of salinity tolerance, maintaining pigment stability at lower salt concentrations but being susceptible at higher levels.

Table 1. Data on chlorophyll a and b levels and total chlorophyll (mg/L)

Sample Name	A ₆₄₉ (Sample absorbance)	A ₆₆₅ (Sample absorbance)	Chl. a content (mg/L)	Chl. b content (mg/L)	Total Chlorophyll (mg/L)
<i>Dendrobium</i> sp. Kontrol	0,16	0,31	3,33	1,69	5,02 ± 2,04 ^b
<i>Dendrobium</i> sp. - NaCl 1%	0,25	0,52	5,75	2,45	8,20 ± 2,04 ^a
<i>Dendrobium</i> sp. - NaCl 5%	0,12	0,24	2,58	1,16	3,74 ± 2,04 ^b
<i>Dendrobium</i> sp. - NaCl 10%	0,12	0,26	2,89	1,15	4,05 ± 2,04 ^b
<i>Phalaenopsis</i> sp. Kontrol	0,17	0,32	3,37	1,88	5,25 ± 2,10 ^a
<i>Phalaenopsis</i> sp. - NaCl 1%	0,12	0,22	2,30	1,48	3,78 ± 2,10 ^b
<i>Phalaenopsis</i> sp. - NaCl 5%	0,05	0,12	1,36	0,38	1,74 ± 2,10 ^c
<i>Phalaenopsis</i> sp. - NaCl 10%	0,02	0,02	0,18	0,36	0,54 ± 2,10 ^d
<i>Vanda</i> sp. Kontrol	0,29	0,58	6,32	3,09	9,42 ± 2,90 ^a
<i>Vanda</i> sp. - NaCl 1%	0,29	0,58	6,28	3,13	9,41 ± 2,90 ^a
<i>Vanda</i> sp. - NaCl 5%	0,14	0,29	3,22	1,31	4,53 ± 2,90 ^b
<i>Vanda</i> sp. - NaCl 10%	0,13	0,27	2,94	1,31	4,26 ± 2,90 ^b

Note: Data are presented as mean ± standard deviation. Different superscript letters in the same column indicate significant differences at $p < 0.05$

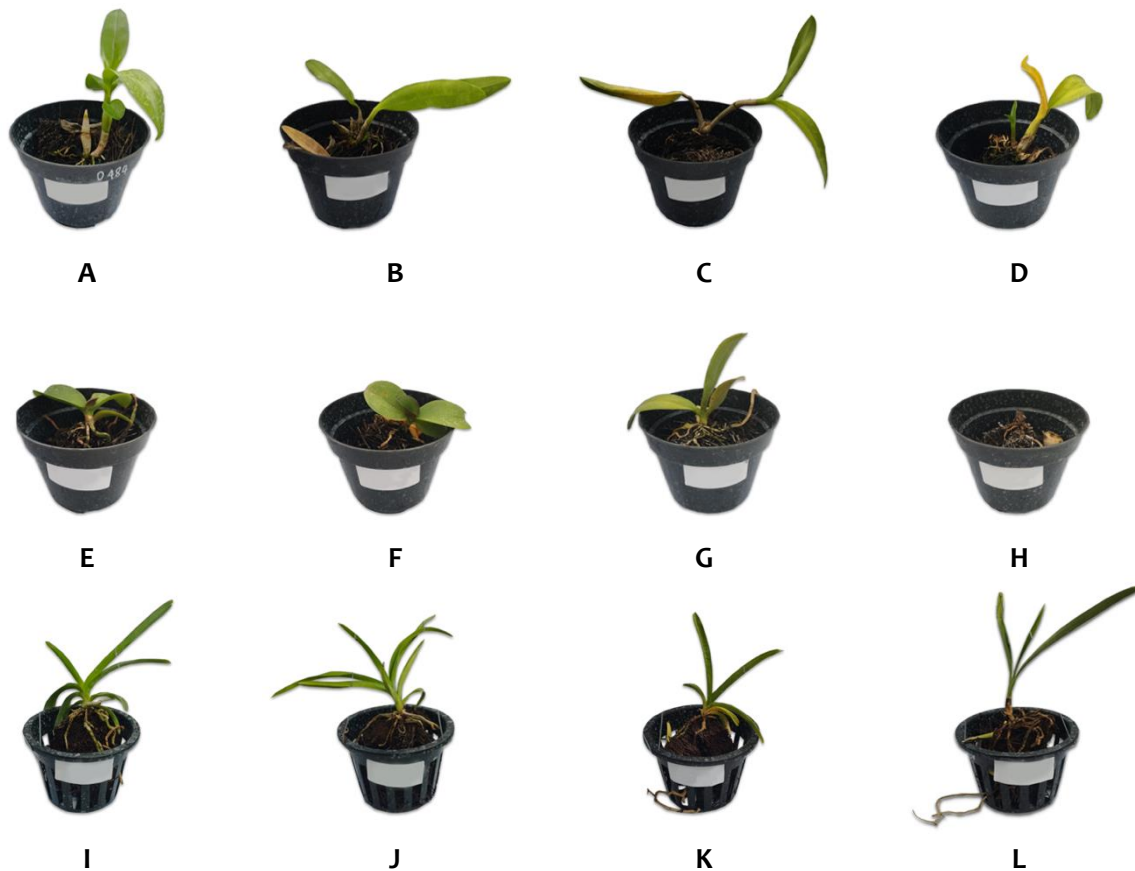


Figure 1. Morphology of the orchid after salinity stressors. *Dendrobium* sp. showed moderate tolerance. (A-D), *Phalaenopsis* sp. was the most sensitive (E-H), and *Vanda* sp. showed the highest tolerance (I-L) treated with 1% NaCl (B,F,J), 5% NaCl (C,G,K), 10% NaCl (D,H,L) compared with the control (A,E,I)

The chlorophyll content of *Vanda* sp. remained virtually unchanged between the control (9.42 mg/L) and 1% NaCl (9.41 mg/L), indicating no negative impact at low salinity. At 5% and 10% NaCl, chlorophyll content decreased to 4.53 mg/L and 4.26 mg/L, respectively, indicating moderate but manageable pigment reduction. Morphological symptoms were relatively mild, consisting of leaf yellowing limited to older leaves without severe necrosis. Compared with other genera, *Vanda* sp. showed the strongest tolerance to salinity. Based on chlorophyll content and morphological response, the order of relative tolerance to salinity was *Vanda* sp. > *Dendrobium* sp. > *Phalaenopsis* sp.

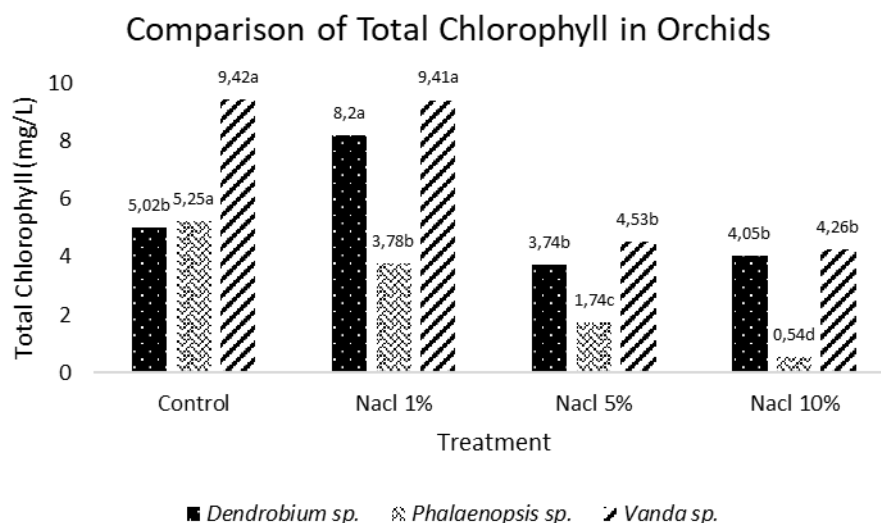


Figure 2. Comparison of total chlorophyll in *Dendrobium sp.*, *Phalaenopsis sp.*, and *Vanda sp.* orchids treated with several different NaCl concentrations

4. Discussion

The results clearly show that salinity stress, caused by NaCl, significantly reduces chlorophyll content and causes morphological damage in orchids, but the tolerance level varies across genera. The drastic decrease in chlorophyll in *Phalaenopsis sp.* indicates its high sensitivity to salt stress. A decrease from 5.25 mg/L (control) to 0.54 mg/L (10% NaCl) is associated with severe chlorosis and necrosis. This indicates that *Phalaenopsis sp.* is unable to regulate ion uptake or maintain osmotic balance under salinity stress. Salinity is known to disrupt chloroplast ultrastructure, increase the accumulation of reactive oxygen species (ROS), and inhibit the chlorophyll biosynthesis pathway [3; 8]. *Phalaenopsis sp.* showed greater sensitivity to salinity stress, which may be associated with less effective physiological adaptation mechanisms compared to more tolerant orchid genera.

In contrast, *Dendrobium sp.* showed an increase in chlorophyll content under 1% NaCl. This response is likely due to hormesis, where low levels of stress stimulate metabolic activity, including increased chlorophyll biosynthesis and antioxidant defenses. Several studies have reported similar findings in epiphytic and halophytic orchids, where mild

stress can activate the production of osmolytes (e.g., proline, glycine betaine) and ROS-scavenging enzymes, thereby enhancing short-term photosynthetic performance [19; 20]. However, at higher salt concentrations (5–10%), the protective mechanisms of *Dendrobium sp.* appear inadequate, leading to chlorophyll degradation and mild necrotic symptoms.

Vanda sp. exhibits the highest tolerance level, maintaining stable chlorophyll levels at low salinities and showing only moderate declines at higher concentrations. Morphological damage is minimal, limited to leaf yellowing in older tissues. This suggests the presence of efficient protective mechanisms, such as compartmentalization of toxic ions within vacuoles, maintenance of membrane integrity, and activation of stress-responsive genes. Recent studies have highlighted that *Vanda* orchids exhibit greater metabolic plasticity, including efficient stomatal regulation and ROS detoxification, compared to other genera [9]. This makes *Vanda sp.* a strong candidate for breeding salt-tolerant orchid cultivars.

The salinity tolerance ranking (*Vanda sp.* > *Dendrobium sp.* > *Phalaenopsis sp.*) highlights the importance of genotypic variation among

orchids. It also supports the idea that orchids, as tropical epiphytes, employ diverse adaptive strategies that reflect their ecological niches. From a practical perspective, identifying tolerant orchids like *Vanda* provides valuable genetic resources for germplasm conservation and breeding programs. Furthermore, understanding the morphophysiological basis of salinity tolerance can inform in vitro selection, genetic engineering, and genome editing approaches aimed at improving abiotic stress tolerance in orchids. Overall, these findings confirm that NaCl salinity is a major limiting factor in orchid cultivation due to its strong inhibitory effect on chlorophyll content and photosynthesis [21]. However, the variation in tolerance between genera highlights opportunities for conservation and improvement. The development of salinity-tolerant orchid varieties will be crucial to ensure sustainable orchid production, especially in areas affected by soil salinization and climate change.

5. Conclusion

This study demonstrates differences in morphophysiological responses among *Dendrobium* sp., *Phalaenopsis* sp., and *Vanda* sp. under NaCl-induced salinity stress. *Phalaenopsis* sp. showed the highest sensitivity, *Dendrobium* sp. exhibited moderate tolerance, and *Vanda* sp. showed the greatest tolerance by maintaining relatively stable chlorophyll levels and minimal morphological damage. These findings highlight variation in salinity resistance among orchid genera and identify *Vanda* sp. as a potential candidate for developing salinity-tolerant varieties, with implications for orchid cultivation and conservation under increasing salinity stress.

6. REFERENCES

- [1] S. W. Gale, G. A. Fischer, P. J. Cribb, and M. F. Fay, "Orchid Conservation: Bridging the Gap between Science and Practice," 2018. [Online]. Available: [https://academic.oup.com/botlinnean/arti](https://academic.oup.com/botlinnean/article-abstract/186/4/425/4955326)
- [2] Nurmalinda, D. Siti Badriah, and S. Kartikaningrum, "Consumer Preference Analysis of *Phalaenopsis* Orchid Variety," in *E3S Web of Conferences*, EDP Sciences, Nov. 2023. doi: 10.1051/e3sconf/202344402004.
- [3] Y. Li, Z. Kang, X. Zhang, P. Sun, X. Jiang, and Z. Han, "The Mycorrhizal Fungi of *Cymbidium* promote the Growth of *Dendrobium officinale* by Increasing Environmental Stress Tolerance," *PeerJ*, vol. 9, Dec. 2021, doi: 10.7717/peerj.12555.
- [4] M. Mustaqim, A. Dan, and A. Listiawati, "Pertumbuhan Anggrek Hitam pada Simulasi Cekaman Kekeringan Menggunakan Polietilen Glikol secara In Vitro Black Orchid Growth in Simulated Drought Stress Using Polyethylene Glycol in Vitro," Dec. 2013.
- [5] N. D. Swarts and K. W. Dixon, "Perspectives on Orchid Conservation in Botanic Gardens," Nov. 2009. doi: 10.1016/j.tplants.2009.07.008.
- [6] R. Soelistijono et al., "Mikoriza Rhizoctonia: Sebagai Agen Ketahanan pada Anggrek *Dendrobium*," vol. 25, p. 89, 2025, doi: 10.36728/afp.v22i2.4612.
- [7] C. Leng, M. Hou, Y. Xing, and J. Chen, "Perspective and Challenges of Mycorrhizal Symbiosis in Orchid Medicinal Plants," Apr. 01, 2024, Elsevier B.V. doi: 10.1016/j.chmed.2024.03.001.
- [8] C. Zhang, J. Chen, W. Huang, X. Song, and J. Niu, "Transcriptomics and Metabolomics Reveal Purine and Phenylpropanoid Metabolism Response to Drought Stress in *Dendrobium sinense*, an Endemic Orchid Species in Hainan Island," *Front. Genet.*, vol. 12, Jul. 2021, doi: 10.3389/fgene.2021.692702.
- [9] J. M. Barreda-Castillo, J. L. Monribot-Villanueva, N. Velázquez-Rosas, P. Bayman, J. A. Guerrero-Analco, and R. A. Menchaca-García, "Morphological and Physio-Chemical Responses to PEG-Induced Water Stress in *Vanilla planifolia*

- and V. pompona Hybrids,” *Int. J. Mol. Sci.*, vol. 24, no. 5, Mar. 2023, doi: 10.3390/ijms24054690.
- [10] L. Hao et al., “The varying responses of leaves and roots and the link between sugar metabolic genes and the SWEET family in *Dendrobium officinale* under salt stress,” *BMC Genomics*, vol. 25, no. 1, Dec. 2024, doi: 10.1186/s12864-024-11069-5.
- [11] S. Mahajan and N. Tuteja, “Cold, Salinity and Drought Stresses: An Overview,” *Arch. Biochem. Biophys.*, vol. 444, no. 2, pp. 139–158, Dec. 2005, doi: 10.1016/j.abb.2005.10.018.
- [12] X. Liu et al., “Effects of Soil Water and Nitrogen Availability on Photosynthesis and Water Use Efficiency of *Robinia pseudoacacia* Seedlings,” *J. Environ. Sci. (China)*, vol. 25, no. 3, pp. 585–595, Mar. 2013, doi: 10.1016/S1001-0742(12)60081-3.
- [13] N. S. Ai and Y. Banyo, “Konsentrasi Klorofil Daun sebagai Indikator Kekurangan Air pada Tanaman,” *Jurnal Ilmiah Sains*, vol. 11, no. 2, pp. 166–173, Oct. 2011, doi: 10.35799/jis.11.2.2011.202.
- [14] I. O. Nabilla and E. Nurcahyani, “Analisis kandungan klorofil pada familia orchidaceae terhadap cekaman kekeringan,” *Cassowary*, vol. 5, no. 2, pp. 134–139, Jun. 2022, doi: 10.30862/cassowary.cs.v5.i2.169.
- [15] M. F. Huda, S. Indriyani, and W. Widoretno, “The Effect of Explant Types and Plant Growth Regulators On Callus Induction of *Geranium (Pelargonium graveolens L’Her)* In Vitro,” in *IOP Conference Series: Earth and Environmental Science*, Institute of Physics Publishing, Dec. 2019. doi: 10.1088/1755-1315/391/1/012034.
- [16] M. F. Huda, O. P. Y. Meishanti, and D. A. Agustin, “Study of the Ethnobotanical Use of Herbal Plants in Blitar District,” *El-Hayah: Jurnal Biologi*, vol. 9, no. 3, pp. 107–115, Sep. 2023, doi: 10.18860/elha.v9i3.28881.
- [17] H. Liu, L. Wang, X. Jing, Y. Chen, and F. Hu, “Functional Analysis of CgWRKY57 from *Cymbidium goeringii* in ABA Response,” *PeerJ*, vol. 9, Feb. 2021, doi: 10.7717/peerj.10982.
- [18] J. F. G. M. Wintermans and A. De Mots, “Spectrophotometric Characteristics of Chlorophylls a and b and Their Pheophytins in Ethanol,” *Biochemica Et Biophysica Acta*, vol. 109, pp. 448–453, 1965.
- [19] S. Tay, J. He, and T. W. Yam, “CAM Plasticity in Epiphytic Tropical Orchid Species Responding to Environmental Stress,” *Bot. Stud.*, vol. 60, no. 1, Dec. 2019, doi: 10.1186/s40529-019-0255-0.
- [20] S. Redondo-Gómez, E. Mateos-Naranjo, M. E. Figueroa, and A. J. Davy, “Salt Stimulation of Growth and Photosynthesis in An Extreme Halophyte, *Arthrocnemum macrostachyum*,” *Plant Biol.*, vol. 12, no. 1, pp. 79–87, Jan. 2010, doi: 10.1111/j.1438-8677.2009.00207.x.
- [21] M. A. Mudhor, P. Dewanti, T. Handoyo, and T. Ratnasari, “Pengaruh Cekaman Kekeringan Terhadap Pertumbuhan dan Produksi Tanaman Padi Hitam Varietas Jeliteng” “Effect of Drought Stress on Growth and Production of Black Rice Plants of Jeliteng Varieties,” *Jurnal Agrikultura*, vol. 2022, no. 3, pp. 247–256, 2022.